MOLECULAR CLONING AND TISSUE SPECIFIC EXPRESSION OF FRUCTOSE 6-PHOSPHATE,2-KINASE:FRUCTOSE 2,6-BISPHOSPHATASE OF RAT BRAIN 1

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A cDNA clone (3591 base pairs) encoding an isozyme of Fructose 6-phosphate, 2-kinase:Fructose 2,6-bisphosphatase was isolated from a rat brain cDNA library. The 5' sequence of this clone (1241 base pairs) was identical to that of the heart type isozyme cDNA (except for 7 base pair mismatches), but the 3' nucleotide sequence (2024 base pairs) was completely different. Its deduced amino acid sequence showed that the enzyme lacked a regulatory domain which contained phosphorylation sites for protein kinase A and C. The results of Northern blotting and polymerase chain reaction demonstrated that the mRNA, 7.4 kilobases long, was expressed also in heart, testis, liver, and skeletal muscle.

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Fructose 2,6-bisphosphate is the most potent activator of phosphofructokinase and plays an important role in regulation of glycolysis (1). Its synthesis and degradation are catalyzed by a bifunctional enzyme, Fructose 6-phosphate,2-kinase:Fructose 2,6-bisphosphatase (Fru 6-P,2-kinase:Fru 2,6-Pase). The intracellular concentration of Fructose 2,6-bisphosphate is determined by the relative activities of the kinase and the phosphatase of the bifunctional enzyme. Thus far, four major isozymes of mammalian enzymes, including liver (2), skeletal muscle (3), heart (4), and testis (5) have been characterized.

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Abbreviations: Fru 6-P 2-kinase: Fru 2,6-Pase, fructose 6-phosphate 2-kinase: fructose 2,6-bisphosphatase; PCR, polymerase chain reaction; bp, base pairs; kb, kilobase pairs.

Recently, Ventura *et al.* (6,7) reported the occurrence of a unique isozyme in bovine and rat brain whose enzyme activities are not affected by protein kinase A or protein kinase C. They isolated an isozyme from bovine brain with a subunit M_r of 120,000 (6), which is $2 \times \text{larger}$ than the other known mammalian bifunctional enzymes.

Our earlier immunological studies² showed the presence of a heart type isozyme with subunit M_r =54,000 in rat brain. In order to characterize the brain-specific Fru 6-P,2-kinase:Fru 2,6-Pase further, molecular cloning of rat brain isozyme(s) was performed. In this paper, we present the identification of a new isozymic form in brain which was a product of a rat heart gene by an alternative splicing process. The results of cDNA and amino acid sequence determinations as well as tissue-specific expression are presented.

EXPERIMENTAL PROCEDURES

<u>Materials</u> - Reagents were obtained as follows: cDNA synthesis kit from Amersham Corporation (Arlington Heights, IL); restriction endonucleases from Toyobo (Tokyo, Japan) and New England BioLabs (Beverly, MA); SuperScript RNase H⁻ reverse transcriptase from Gibco-BRL (Gaithersburg, MD), Taq DNA polymerase from Perkin-Elmer Cetus Instruments (Norwalk, CT).

Synthesis and PCR Amplification of cDNA - The oligo(dT)-primed cDNA was synthesized from poly(A)⁺-RNA (1 μ g), using 200 units SuperScript RNaseH reverse transcriptase in 20 μ l of reaction mixture according to the manufacturer's instructions. An aliquot (1 μ l) of the reaction mixture was amplified in 100 μ l of 10 mM Tris-HCl buffer (pH 8.3), containing 50 mM KCl, 1.5 mM MgCl₂, 0.001% (w/v) gelatin, 0.2 mM of each deoxyribonucleotide, 2.5 unit Taq DNA polymerase, and 1 μ M oligonucleotide primers, 5'-CTCCACATCATCTTCAA-3', corresponding to #1447-1463 of RB10 cDNA and 5'-TGACAGCGTCAACAGGC-3', corresponding to #2512-2528 (8). The polymerase chain reaction (PCR) consisted of 30 cycles of denaturation at 94°C for 30 sec, primer annealing 53°C for 20 sec, and primer extension at 72°C for 1 min, with an additional 2 min primer extension included after the final cycle.

RESULTS AND DISCUSSION

Molecular Cloning of Fru 6-P 2-kinase:Fru 2,6-Pase cDNA from Rat Brain - cDNA was synthesized with oligo(dT) primer using rat brain poly(A)⁺-RNA (5 μ g) as a template. The cDNA library containing 3×10^6 independent clones was constructed using λ gt10 phage vector and screened using as probes Fru 6-P,2-kinase:Fru 2,6-Pase cDNAs from bovine heart, human liver, and frog skeletal muscle (manuscript in preparation). Several

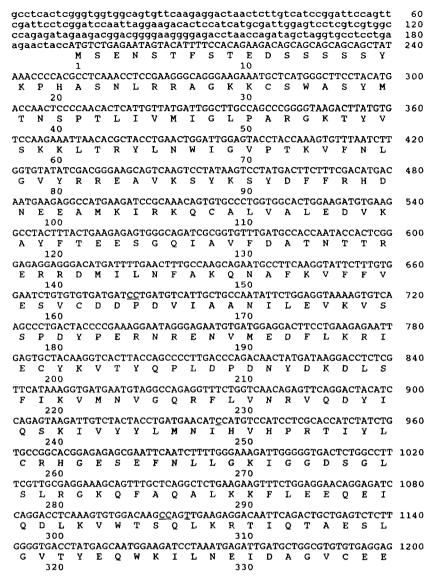
²Unpublished results of R. Sakakibara, M. Taniyama, and K. Uyeda.

screenings yielded five positive clones which harbored cDNA inserts ranging from 0.5 kilobase pairs (kb) to 3.4 kb. Four of these clones were found to share common DNA sequences by restriction enzyme mapping and partial nucleotide sequencing. RB10 containing the 3.4 kb insert was chosen for complete analysis. The fifth clone, RB7, contained a 0.9 kb insert that was not identical by restriction mapping to the other clones. None of the clones contained a large enough insert to encode the enzyme similar to that reported by Ventura *et al.* (6).

cDNA Sequence and Deduced Amino Acid Sequence of Rat Brain Fru 6-P, 2-kinase:Fru 2,6-Pase - The 5' region (308-1548 bp) of RB10 was identical to that of one of the heart isozyme cDNAs previously reported (5) except for 7 base mismatches (Fig. 1, underlined). This common 5' region includes the well-conserved catalytic domains and corresponds to a genomic sequence derived from rat heart exon 3 to exon 14 (9). It seems likely that this clone (derived from Sprague-Dawley rats in Japan) and the rat heart isozyme (derived from S.D. rats in US) represent the same gene, and the 7 base mismatches are probably due to differences in rat sources. However, there was no homology between the 3' regions (1549-3572 bp) of the clone and the rat heart cDNA. This disparate region of the rat heart cDNA is derived from exon 15 and 16 of the gene (9), and it encodes a polypeptide containing Ser and Thr which are known to be phosphorylated by cAMP-dependent protein kinase and protein kinase C (10). The nucleotide sequence analysis of the other three clones revealed that they also lacked this peptide. The 3' sequence of RB10 showed high homology with exon 16 of the bovine heart isozyme gene (manuscripts in preparation), but not with exon 16 of the rat heart gene (9). These results suggest that RB10 may arise by a tissue-specific alternative splicing process using an as yet unidentified exon of the rat heart gene.

The fifth clone, RB7, did not harbor a full length cDNA but contained only a coding region whose deduced amino acid sequence was 70-80% identical to the known mammalian bifunctional isozymes, including heart. These results suggested the existence of multiple isozymes of Fru 6-P,2-kinase:Fru 2,6-Pase in rat brain.

Northern Blotting of RB10 mRNA - To study the *in vivo* expression of RB10 DNA. Northern blotting hybridization was carried out with poly(A)⁺-RNA isolated from various rat tissues. As shown in Figure 2, a single intense signal corresponding to 7.4 kb was clearly detected with mRNA from brain, heart, testis, and liver. The intensity of these bands was approximately the same, but that of skeletal muscle was too weak to be



<u>Figure 1.</u> cDNA sequence of RB10 and its deduced amino acid sequence. Nucleotides are numbered in the 5' to 3' direction. The deduced amino acid residues are given in one-letter code and are indicated below the nucleotide triplets. Nucleotides representing the proposed coding region are capitalized. The nucleotides of 5'-region (308-1548 bp) of RB10 which show mismatch with rat heart cDNA are underlined.

shown in an autoradiogram. However, the same band was detected with skeletal muscle upon longer exposure. Ventura *et al.* (7) also reported a similar band at 6.8 kb in rat brain and heart using the rat liver bifunctional enzyme cDNA as a probe.

<u>Tissue Specific Alternative Splicing of RB10</u> - In order to determine whether the mRNA that encodes the peptide containing the phosphorylation sites of the heart type enzyme

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ATGACTTATTCGGAGATC\underline{Q}AACAACGGTATCCAGAGGAATTTGCACTTCGAGATCAAGAG 1260 M T Y S E I E Q R Y P E E F A L R D Q E
                               350
AAGTATCTGTATCGATATCCTGGTGGGGAGTCATACCAGGACCTGGTGCAGCGGCTGGAG 1320
   YLYRYPGGES
                             YQDLVQRLE
                               370
CCTGTGATCATGGAGCTGGAGCGCAAGGCAACGTCCTCGTTATCTCTCACCAGGCTGTC 1380
     I M E L E R Q G N V L V I S H Q A
    380
                              390
ATGCGCTGCCTCCTGGCCTACTTCTTGGATAAAGGCGCAGATGAGTTGCCGTACTTGAGG 1440
   DELPYL
    400
                              410
TGCCCCCTCCACATCATCTTCAAACTTACTCCTGTGGCCTATGGTTGCAAAGTGGAAACA 1500
  P L H I I F K L T P V A
420 430
                                  YGCKV
ATTACACTGAATGTGGAAGCTGTGGACACACATCGTGACAAGCCAACTGAAGTGGAGAAT 1560
   T L N V E A V D T H R D
                                  KPTEV
    440
                              450
GTGCTGGCTAAGCATAGACGCCCCTCAATGGCATCCCTCACTTTGCTCTCCtgatqtqag 1620
        KHRRPSMAS
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gctgaggccagacctctccgaggaactggatctaccaaaaagcttggcatgccagcatcc
ctgaggctagaacaggaagttaagtgggaacgtctcatttgaggctaccaacagtaaggc 1740
gtgggaaaccttcactgcatttctacccctcatgcccaacaggaacattactacattgtg 1800
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                                                     2100
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                                                     2160
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                                                     2220
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tacattacatatgcacaaaatattaacattttccaaaatgtacttgaaagtttattaagg 3540
3591
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Figure 1 - Continued

occurs in other rat tissue, mRNAs from brain, testis, liver, and skeletal muscle were analyzed by PCR. The cDNAs synthesized from those mRNAs were amplified with synthetic oligonucleotide primers corresponding to #1447-1463 and to #2512-2528 nucleotide residues of RB10. The PCR amplification detected a band (1.1 kb) in all lanes, the size expected if derived from mRNAs which lack sequences encoding the phosphorylation domain of the enzyme in these tissues (Fig. 3). An additional band of 1.3 kb was detected in rat heart and skeletal muscle (lane 2 and lane 5 in Fig. 3). The additional 0.2 kb DNA of the 1.3 kb band may correspond to the phosphorylation site peptide.

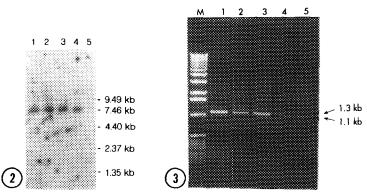


Figure 2. Tissue-specific expression of Fru 6-P,2-kinase:Fru 2,6-Pase mRNA in rat tissues. Poly(A)⁺-RNA (10 μ g) from various rat tissues was fractionated by electrophoresis and hybridized with ³²P-labeled RB10 cDNA. The origins of mRNA were as follows: 1, brain; 2, heart; 3, testis; 4, liver; and 5, skeletal muscle.

<u>Figure 3.</u> Analysis of RB10 mRNA by PCR. The synthesis of cDNA and PCR amplification were carried out as described in **EXPERIMENTAL PROCEDURES**. The reaction mixture (10 μ I) was subjected to 1% agarose gel electrophoresis. The origins of mRNA were as follows: 1, brain; 2, heart; 3, testis; 4, liver; and 5, skeletal muscle; M, molecular size standard.

Thus, we conclude that the heart type isozyme of Fru 6-P,2-kinase:Fru 2,6-Pase lacking the phosphorylation domain peptide occurs in rat brain, testis, and liver. The enzyme in these tissues is not responsive to regulation by cAMP. Rat heart and skeletal muscle contain both types of the heart bifunctional isozyme, one of which is subject to regulation by cAMP. Furthermore, the heart type isozyme appears to be the most abundant form among the isozymes in rat brain. We were unable to detect any additional brain-specific isozyme such as that reported by others (6).

REFERENCES

- 1. Uyeda, K., Furuya, E., Richards, C.S., and Yokoyama, M. (1982) Mol. Cell Biochem. 48:97-120
- 2. Sakakibara, R., Kitajima, S., and Uyeda, K. (1984) J. Biol. Chem. 259:41-46
- 3. van Schaftingen, E. and Hers, H.G. (1986) Eur. J. Biochem. 159:359-365
- 4. Kitamura, K. and Uyeda, K. (1988) J. Biol. Chem. 263:9027-9033
- 5. Sakata, J., Abe, Y., and Uyeda, K. (1991) J. Biol. Chem. 266:15764-15770
- Ventura, F., Rosa, J.L., Ambrosio, S., Pilkis, S.J., and Bartrons, R. (1992) J. Biol. Chem. 267:17939-17943
- Ventura, F., Rosa J.L., Ambrosio, S., Gil, J., and Bartrons, R. (1991) Biochem. J. 276:455-460
- 8. Saiki, R.K., Gelfand, D.H., Stoffel, S., Scharf, S.J., Higuchi, R., Horn, G.T., Mullis, K.B., and Erlich, H.A. (1988) Science, 239:487-491
- 9. Darville M.I., Chikri, M., Lebeau, E., Hue, L., and Rousseau, G.G. (1991) FEBS Lett. 288:91-94
- Kitamura, K., Kangawa, K., Matsuo, H., and Uyeda, K. (1988) J. Biol. Chem. 263:16796-16801